

# Multivalent neoglycoconjugates: solid-phase synthesis of N-linked $\alpha$ -sialodendrimers

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A series of N-linked  $\alpha$ -sialodendrimers with valencies of 2, 4 and 8 has been scaffolded on an *N,N*-bis(3-aminopropyl)succinamic acid core using Wang resin and 9-fluorenylmethoxycarbonyl and 2-(1*H*-benzotriazol-1-yl)-1,1,3,3-tetramethyluronium tetrafluoroborate solid-phase chemistry.

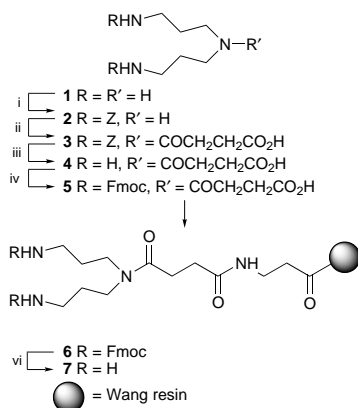
Dendrimers are attractive monodispersed macromolecules with broad spectrum applications from physics to medicine.<sup>1</sup> The design of novel dendrimers represents an active research area and their syntheses are considered challenging owing to their size and highly branched structures. Traditionally, divergent<sup>2</sup> and convergent<sup>3</sup> strategies have been used to synthesize dendrimers. However, more recently a double exponential growth strategy<sup>4</sup> and a self-assembling method<sup>5</sup> have been developed. To date, the synthesis of most dendrimers has been carried out in solution, although Tam<sup>6</sup> has built hyperbranched L-lysine dendrimers on a solid-phase for multiple antigen peptide (MAP) presentation. Because of the critical role of carbohydrate-containing macromolecules in many cellular processes, glycodendrimers have also become a growing research area.<sup>7</sup> There is increasing evidence that the multivalent effect displayed by these novel biopolymers may overcome the characteristic low-affinity binding interactions between carbohydrate ligands and proteins.<sup>8,9</sup>

Potent L-lysine-based dendritic  $\alpha$ -thiosialoside inhibitors of *Influenza* virus's hemagglutination to human erythrocytes have been previously described.<sup>10</sup> Similarly, different series of symmetrical dendrimers containing *N*-acetylneuraminic acid (NeuAc or sialic acid), representing one of the most widespread mammalian cell surface carbohydrate ligands,<sup>11</sup> have been successfully synthesized in solution.<sup>12,13</sup> Therefore, it was attractive to build a new series of sialodendrimers on a solid-phase. The present approach differs from previous S-linked sialosides<sup>10,11</sup> by the use of N-linked sialylated derivatives which are also expected to be sialidase resistant. Thus, the solid-

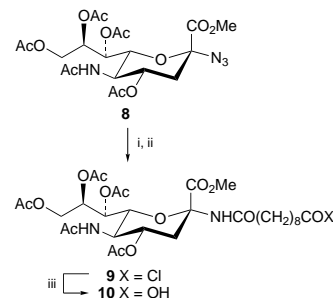
phase synthesis of a new family of symmetrical N-linked sialodendrimers based on *N,N*-bis(3-aminopropyl)succinamic acid **4** core is described.

In order to provide a strategy which could be used with other carbohydrate ligands, a divergent approach was used to build these novel poly(amidoamine) dendrimers on a solid-phase. Suitably acid-functionalized carbohydrate derivatives could then be added onto polyamino dendrimers to afford glycodendrimers with even valencies. The dendritic core was prepared as follows (Scheme 1). The primary amine groups of *N*-(3-aminopropyl)propane-1,3-diamine **1** were regioselectively protected with benzyloxycarbonyl (Z) groups using benzyl cyanofornate in  $\text{CH}_2\text{Cl}_2$ <sup>14</sup> to give secondary amine **2** in 72% yield. Acylation of the residual amine with succinic anhydride (triethylamine,  $\text{CH}_2\text{Cl}_2$ , 96%) provided key building block **3** having a divalent structure with a bis-Z-protected monoacid core. In order to use the Fmoc strategy, the Z-protecting groups were removed by catalytic hydrogenation (10% Pd-C, MeOH) to give **4**, which was then treated with 9-fluorenylmethyl chloroformate (FmocCl, 10%  $\text{Na}_2\text{CO}_3$ -dioxane, 0 °C) to afford **5**<sup>†</sup> in 72% yield. The dendritic polyamine scaffolds were built on a  $\beta$ -alanine spacer attached to Wang resin [4-(hydroxymethyl)phenoxymethyl-*co*-poly(styrene-1% divinylbenzene), 0.58 mmol g<sup>-1</sup>], using the Fmoc-strategy and performing the deprotection-coupling cycles as follow: (i) Fmoc deprotection by treatment with 20% piperidine in DMF (1  $\times$  5 min, 1  $\times$  15 min), (ii) coupling of Fmoc-protected acid **5** (2 equiv.) using 2-(1*H*-benzotriazol-1-yl)-1,1,3,3-tetramethyluronium tetrafluoroborate (TBTU) (2 equiv.) as activating agent and  $\text{Pr}^i_2\text{EtN}$  (4 equiv.) in DMF during 30 min. Coupling completion was determined by a ninhydrin test for residual amino groups.<sup>15</sup> To characterize the dendrimers, 30 mg of resin from each sequential generation was Fmoc-deprotected and the dendrimers were released from the solid support following conventional acid treatment (95% aq. TFA, 2 h, 30–79% yield).

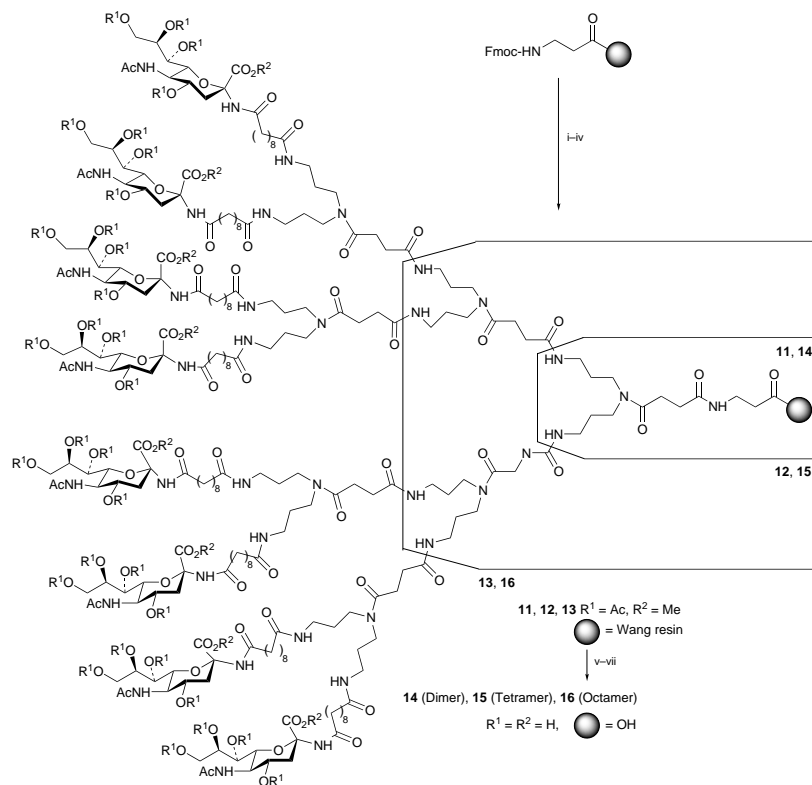
Glycodendrimer synthesis required sialic acid residues suitably functionalized at the anomeric position for coupling with amine-terminated dendrimers. The readily available  $\alpha$ -sialic acid azide derivative **8**<sup>16</sup> was reduced by catalytic hydrogenation (10% Pd-C, MeOH) to provide  $\alpha$ -sialic acid amine in quantitative yield (Scheme 2). To minimize anomericisation, the next acylation step was performed within 1 h. To



**Scheme 1** Reagents and conditions: i,  $\text{BnO}_2\text{CCN}$ ,  $\text{CH}_2\text{Cl}_2$ , 2 h, 72%; ii, succinic anhydride,  $\text{Et}_3\text{N}$ ,  $\text{CH}_2\text{Cl}_2$ , 1 h, 96%; iii,  $\text{H}_2$ , 10% Pd-C, MeOH, 2 h, quant.; iv, FmocCl, 10%  $\text{Na}_2\text{CO}_3$ -dioxane, 12 h, 72%; v, H- $\beta$ -Ala-Wang resin, TBTU,  $\text{Pr}^i_2\text{EtN}$ , DMF, 30 min; vi, 20% piperidine-DMF (1  $\times$  5 min, 1  $\times$  15 min)



**Scheme 2** Reagents and conditions: i,  $\text{H}_2$ , 10% Pd-C, MeOH, 1 h, quant.; ii,  $\text{ClCO}(\text{CH}_2)_8\text{COCl}$ ,  $\text{Pr}^i_2\text{EtN}$ ,  $\text{CH}_2\text{Cl}_2$ , 3 h; iii,  $\text{H}_2\text{O}$ , 53% overall yield



**Scheme 3** Reagents and conditions: i, 20% piperidine–DMF (1 × 5 min, 1 × 15 min); ii, **5**, TBUTU,  $\text{Pr}_2\text{EtN}$ , DMF, 30 min; iii, repeat cycle or 20% piperidine–DMF; iv, **10**, TBUTU,  $\text{Pr}_2\text{EtN}$ , DMF, 0.5–8 h; v, 95% aq. TFA, 2 h; vi, 1 M NaOMe, MeOH, 2–8 h; vii, 0.05 M NaOH, 2–8 h

offer suitable carbohydrate accessibility, introduction of a spacer arm between the sialic acid residues and the dendrimer was carried out using excess  $\text{ClCO}(\text{CH}_2)_8\text{COCl}$  ( $\text{CH}_2\text{Cl}_2$ ,  $\text{Pr}_2\text{EtN}$ , 0 °C) to give unstable monoacid chloride **9** which was immediately hydrolysed to afford **10** in 53% overall yield. Attempts to isolate **9** for subsequent attachment to the resin were unsuccessful.  $\alpha$ -Sialic acid derivative **10** was then ready to be coupled to the dendritic polyamine core on the solid-phase using the same TBUTU coupling strategy described to scaffold the dendrimer generations. After coupling, peracetylated sialodendrimers with the valencies of 2, 4 and 8 (**11**, **12** and **13**) were released from the resin by treatment with 95% TFA. The sialodendrimers were then deprotected by sequential ester hydrolysis [(i) de-*O*-acetylation with 1 M NaOMe in MeOH, (ii) 0.005 M NaOH]. Purification of each independent generation by size exclusion chromatography over Biogel-P2 ( $\text{H}_2\text{O}$  as eluent) provided pure deprotected glycodendrimers **14**, **15** and **16** in moderate yields (25–56%) (Scheme 3). The purity of each compound was readily established from the relative integration of key signals in the  $^1\text{H}$  NMR spectrum. MALDI-TOF mass spectral data (negative mode) further confirmed the integrity of the dendrimers. Biological properties of these novel N-linked sialodendrimers will be reported in due course.

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## Footnotes and References

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† All compounds showed consistent NMR and mass spectral data. Because of dendrimers' repetitive structures, only selected data are reported. Selected data for **5**:  $\delta_{\text{H}}([\text{D}_6]\text{acetone})$  1.69 and 1.91 (2 m, 4 H,  $\beta\text{-CH}_2$ ), 2.6 [m, 2 H, succinyl  $\text{CH}_2\text{C}(\text{O})\text{OH}$ ], 4.3 (dd, 4 H, Fmoc- $\text{CH}_2$ s);  $\delta_{\text{C}}$  28.7 and 30.5 ( $\beta\text{-C}$ ), 29.8 [succinyl  $\text{CH}_2\text{C}(\text{O})\text{OH}$ ]; FAB-MS (positive): calc. for  $\text{C}_{40}\text{H}_{41}\text{N}_5\text{O}_7$ : 675.7. Found: 676.3 ( $\text{M} + 1$ ). For **10** (spacer identification from anomeric position to acid is a to h):  $\delta_{\text{H}}(\text{CDCl}_3)$  1.59 (m, 4 H, b-,

g- $\text{CH}_2$ s), 1.87 (s, 3 H, NAc), 2.07 (m, 1 H, H-3 $^{\text{ax}}$ ), 2.74 (m, 1 H, H-3 $^{\text{eq}}$ ), 3.74 (s, 3 H,  $\text{OCH}_3$ ), 5.35 (dd, 1 H,  $J_{6,7}$  2.2,  $J_{7,8}$  6.1, H-7);  $\delta_{\text{C}}$  23.14 (NAc), 24.58 (g-C), 52.9 ( $\text{OCH}_3$ ), 83.3 (C-2); FAB-MS (positive): calc. for  $\text{C}_{30}\text{H}_{46}\text{N}_2\text{O}_{15}$ : 674.7. Found: 675.4 ( $\text{M} + 1$ ). For **14**:  $\delta_{\text{H}}(\text{D}_2\text{O})$  1.62 (m, b-, g- $\text{CH}_2$ s), 1.84 (m, 6 H,  $\beta\text{-CH}_2$ , H-3 $^{\text{ax}}$ ), 2.08 (s, 6 H, NAc), 2.44 (m, 2 H,  $\beta$ -alanyl  $\alpha\text{-CH}_2$ ), 2.73 [m, 4 H, succinyl  $\text{CH}_2\text{C}(\text{O})\text{N}$ , H-3 $^{\text{eq}}$ ], 4.16 (d, 2 H,  $J$  9.9, H-7);  $\delta_{\text{C}}$  21.5 (NAc), 24.5 (g-C), 26 and 27 ( $\beta\text{-Cs}$ ), 36.2 ( $\gamma\text{-Cs}$ ,  $\beta$  alanyl  $\alpha\text{-C}$  and  $\beta\text{-C}$ ), 72.4 (C-7), 84.4 (C-2); FAB-MS (positive): calc. for  $\text{C}_{55}\text{H}_{94}\text{N}_8\text{O}_{24}$ : 1250.64. Found: 1251.5 ( $\text{M}^+ + 1$ , 1%); MALDI-TOF (negative): Found: 1250.03 ( $\text{M} - 1$ ) $^-$ . For **15**: MALDI-TOF (negative): calc. for  $\text{C}_{117}\text{H}_{200}\text{N}_{18}\text{O}_{48}$ : 2625.37. Found: 2626 ( $\text{M} - 1$ ) $^-$ . For **16**:  $\delta_{\text{H}}(\text{D}_2\text{O})$  1.86 (m, 28 H,  $\beta\text{-CH}_2$ s), 2.11 (s, 24 H, NAc), 4.18 (m, 8 H, H-7);  $\delta_{\text{C}}$  21.57 (NAc), 26.2 ( $\beta\text{-Cs}$ ), 72.5 (C-7), 85.2 (C-2); MALDI-TOF (negative) calc. for  $\text{C}_{241}\text{H}_{412}\text{N}_{38}\text{O}_{96}$ : 5374.85. Found: 5374 ( $\text{M} - 1$ ) $^-$ .

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